

Fluidigm System for Rapid, Reliable, and Cost Effective SNP Genotyping

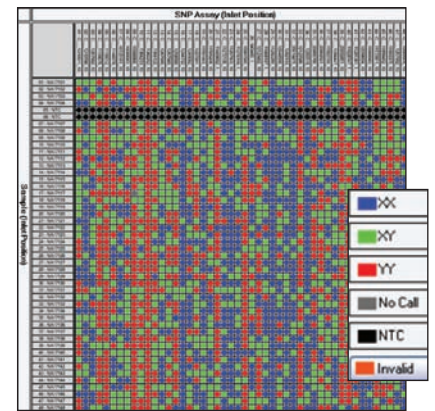
Single nucleotide polymorphisms (SNPs) are the most common form of genetic variation found in the genome of any organism. Although SNP genotypes are a powerful tool for understanding disease disposition and drug metabolism, SNP genotyping of large populations has been impractical because of the running costs and time associated with conventional microwell plate-based systems.

Comparatively, the Fluidigm system, including Fluidigm 48.48 Dynamic Arrays, is a nanofluidic system that is far more efficient while yielding equivalent call rates and reproducibility. Instead of robotic liquid-handling required to setup microwell plates, dynamic arrays have an integrated network of channels, chambers, and valves that automatically combine the reactions, conserving time, reagent, and sample amount. Following are results of experiments designed to validate call reproducibility of the Fluidigm system — and to demonstrate SNP genotyping concordance with the ABI 7900.

SNP Genotyping

Overview. Forty-eight TaqMan® SNP genotyping assays were obtained from Applied Biosystems, Foster City, CA (Table 1). Five microliters of 10x assays were loaded into the primer/probe inlets of two dynamic arrays. Forty-six different DNA samples (159–186 ng/μl) were obtained from Coriell Cell Repositories, Camden, NJ (Table 2). Each sample was mixed with 2x Universal TaqMan® PCR Master Mix (Applied Biosystems) and loaded into the sample inlets of the dynamic arrays.

Results. The results are displayed as an allele map, with each data point a pair-wise combination of a specific SNP primer/probe and sample. The results show call rates greater than 99.5% and high concordance between the duplicate dynamic array runs.



Assays Used

| # | Assay Name | # | Assay Name | # | Assay Name |
|----|---------------|----|---------------|----|---------------|
| 1 | C_1210911_10 | 2 | C_1210596_10 | 3 | C_1207992_10 |
| 4 | C_1159676_10 | 5 | C_1156012_10 | 6 | C_1146078_10 |
| 7 | C_16125161_10 | 8 | C_16085063_10 | 9 | C_15988806_10 |
| 10 | C_1364233_10 | 11 | C_1382252_10 | 12 | C_1364122_10 |
| 13 | C_1267995_10 | 14 | C_1267960_10 | 15 | C_1258500_10 |
| 16 | C_1244732_10 | 17 | C_1243062_10 | 18 | C_1227201_10 |
| 19 | C_11561443_10 | 20 | C_11505408_10 | 21 | C_11469692_10 |
| 22 | C_11272136_10 | 23 | C_11207933_10 | 24 | C_11276169_10 |
| 25 | C_15961560_10 | 26 | C_940460_10 | 27 | C_12123462_10 |
| 28 | C_11668187_10 | 29 | C_11282289_10 | 30 | C_12048556_10 |
| 31 | C_1282193_10 | 32 | C_25618415_10 | 33 | C_12122586_10 |
| 34 | C_1295519_10 | 35 | C_25800766_10 | 36 | C_25802021_10 |
| 37 | C_11562154_10 | 38 | C_1284447_10 | 39 | C_25650851_10 |
| 40 | C_11628747_10 | 41 | C_11281361_10 | 42 | C_12098654_10 |
| 43 | C_15969983_10 | 44 | C_11572811_10 | 45 | C_11899452_10 |
| 46 | C_11993375_10 | 47 | C_1207992_10 | 48 | C_12005976_10 |

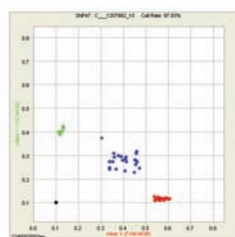
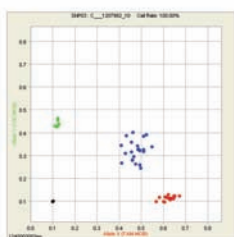
Table 1. SNP genotyping assays used in the study.

| # | Sample ID | # | Sample ID | # | Sample ID |
|----|-----------|----|-----------|----|-----------|
| 1 | NA17101 | 2 | NA17102 | 3 | NA17103 |
| 4 | NA17104 | 5 | NA17105 | 6 | NTC |
| 7 | NA17107 | 8 | NA17108 | 9 | NA17109 |
| 10 | NA17110 | 11 | NA17111 | 12 | NA17316 |
| 13 | NA17113 | 14 | NA17114 | 15 | NA17115 |
| 16 | NA17116 | 17 | NA17117 | 18 | NA17118 |
| 19 | NA1719 | 20 | NA17120 | 21 | NA17121 |
| 22 | NTC | 23 | NA17123 | 24 | NA17124 |
| 25 | NA17125 | 26 | NA17126 | 27 | NA17127 |
| 28 | NA17128 | 29 | NA17129 | 30 | NA17130 |
| 31 | NA17131 | 32 | NA17132 | 33 | NA17133 |
| 34 | NA17134 | 35 | NA17135 | 36 | NA17136 |
| 37 | NA17137 | 38 | NA17138 | 39 | NA17139 |
| 40 | NA17140 | 41 | NA17141 | 42 | NA17142 |
| 43 | NA17143 | 44 | NA17144 | 45 | NA17145 |
| 46 | NA17146 | 47 | NA17147 | 48 | NA17148 |

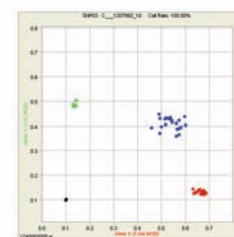
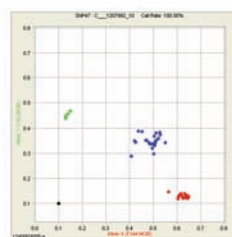
Table 2. DNA samples used in the study.

Reproducibility

Intra- (Panel A or Panel B) and inter-chip (Panel A and B) reproducibility of assay clusters.



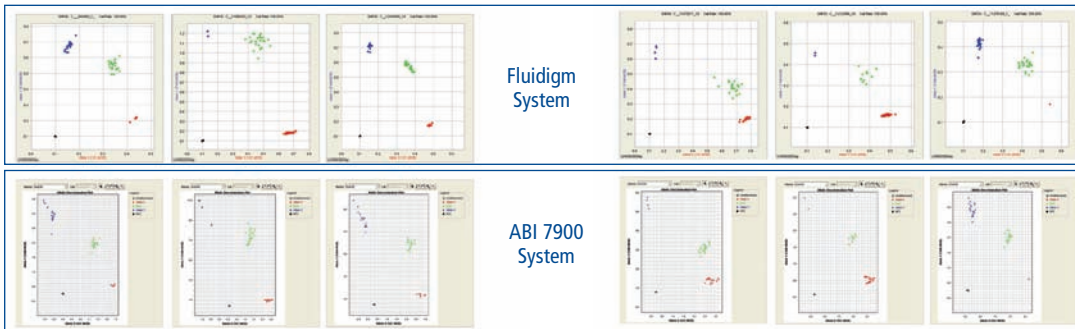
Panel A.



Panel B.

Concordance with Conventional Platforms

The same forty six samples were run against the same 12 primer/probe sets on both the Fluidigm system and a 7900HT Sequence Detection System (Applied Biosystems). Side-by-side comparison shows complete concordance, with a 99.5 percent call rate.



Benefits

Flexibility.

User choice of any 48 samples and any 48 SNP genotyping assays.

Set up and Run time.

Takes less than three hours.

Automation Compatibility.

Standard 384 well format.

Throughput.

Each dynamic array delivers the throughput of six 384-well plates.

SNP Genotyping Analysis Software.

- Auto-calling of SNP genotypes from the clusters.
- Visual overlay of clusters from multiple dynamic arrays.
- Flexible annotation of sample and assay information from the source plate.
- Pipetting maps of samples and primer/probes from the 96-well plates.
- Summary reports, containing the number of clusters, call rates, and confidence for each assay.

Reaction Volumes

Dynamic arrays require much smaller volumes of reaction components, which substantially reduces running costs.

| | 48.48 DYNAMIC ARRAY | SIX 384 WELL PLATE* | FOLD REDUCTION |
|---------------------------------|---------------------|---------------------|----------------|
| 10X ASSAY | 192 μ l | 300 μ l | 1.6X |
| DNA AMOUNT | 150 ng | 480 ng | 5.2X |
| TAQMAN UNIVERSAL PCR MASTER MIX | 150 μ l | 9200 μ l | 61X |

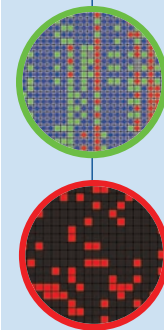
* Assumes reaction volume of 8 μ l

Conclusion

The Fluidigm system is a highly efficient alternative to microwell plate-based systems for SNP genotyping studies requiring 16 or more genes to be screened against 1,000s of samples. The advantages include lower running costs, exponentially higher throughput per run, and simplified setup of reactions. The Fluidigm system provides these advantages while yielding excellent call rates and concordance with widely used microwell plate-based systems.

WORK FLOW

- 1 Prime**
Prime the IFC to prepare for samples and assays.
- 2 Transfer**
Transfer samples and assays into separate inlets on the chip.
- 3 Load**
Place the IFC on the IFC controller to automatically setup reaction chambers.
- 4 Thermal Cycle**
Place the IFC onto the Stand-Alone Thermal Cycler and start the PCR protocol.
- 5 Read**
Place the IFC on the EP1 Reader for fluorescence detection.



Fluidigm

Corporate Headquarters

Fluidigm Corporation
7000 Shoreline Court, Suite 100
South San Francisco, CA 94080 USA
Toll-free: 1.866.FLUIDLINE (1.866.358.4354)
Fax: 650.871.7152
www.fluidigm.com

Sales

North America: 650.266.6170 | biomark@fluidigm.com
Europe/EMEA: +31 20 578 8853 | biomark@fluidigm.com
Japan/Korea: +81 3 3555 2351 | biomarkasia@fluidigm.com
Asia: +65 9431 3790 | biomarkasia@fluidigm.com

© Fluidigm Corporation. All rights reserved.

Fluidigm, the Fluidigm logo, BioMark, EP1, SlingShot, and FLUIDLINE are trademarks or registered trademarks of Fluidigm Corporation in the U.S. and/or other countries. All other trademarks are the property of their respective owners.

Fluidigm recommends that you only purchase TaqMan® dual-labeled probes and/or other licensed PCR assay reagents from authorized sources.

FOR RESEARCH USE ONLY.
MRKT00076e